Application of Molecular Pathology in Endocrine Pathology

Ebru SERİNSOZ LINKE¹, Gaye GÜLER TEZEL²

Department of Pathology, ¹Mersin University Faculty of Medicine, MERSİN, TURKEY, ²Hacettepe University Faculty of Medicine, ANKARA, TURKEY

ABSTRACT

Rapid growth in knowledge of cell and molecular biology led to the increased usage of molecular techniques in anatomical pathology. This is also due to the advances achieved in the techniques introduced in the last few years which are less laborious as compared to the techniques used at the beginning of the “molecular era”. The initial assays were also very expensive and were not performed except for selected centers. Moreover, the clinicians were not sure how to make use of the accumulating molecular information. That situation has also changed and molecular techniques are being performed in a wide variety of medical settings which also has a reflection on the endocrine system pathology among other organ systems. This review will provide an update of genetic changes observed in different endocrine system pathologies and their diagnostic, therapeutic and prognostic values.

Key Words: BRAF mutation, RET/PTC rearrangement, Thyroid, Adrenal, Genetics

INTRODUCTION

Oncogenesis is a complex process with a variety of players involved. The mechanisms underlying this process in different organ systems have been, to some extent, uncovered. Changes at the molecular level, including DNA, RNA and protein changes as well as alterations in small RNA molecules contribute to this complex process. Furthermore external, nutritional, environmental factors, also contribute to carcinogenesis in many organ systems (1).

Tumors in general are subcategorized according to one fundamental feature into either being sporadic or hereditary. In the sporadic tumors the driving changes occur in terminally differentiated cells whereas in hereditary tumors the genetic changes are inherited at the germline level. Sporadic tumors have more complex pathways involved in tumorigenesis as compared to hereditary tumors. The most important contributors of carcinogenesis are tumor associated genes; i.e. tumor suppressor genes and oncogenes. In each normal cell, there are 2 copies of tumor suppressor genes, one inherited from each parent. During tumorigenesis, an inactivating mutation occurs in one copy of the tumor suppressor gene. This usually is called the “first hit”. The first hit can be a sporadic event or can be inherited as is the case in cancer syndromes. The loss of function requires silencing of both copies. Proto-oncogenes are nonmutated genes, which stimulate carcinogenesis when mutated. Overexpression or activation can be achieved only with one mutated copy of the gene as opposed to tumor suppressor genes (1).

Detection of mutation in a tumor suppressor gene can be achieved by loss of heterozygosity analysis. When a mutation occurs in an oncogene, the protein product is usually overexpressed which enables detection either by immunohistochemistry or by molecular techniques which measure mRNA levels (1, 2).

THYROID NEOPLASMS

Thyroid carcinoma is rare among overall human malignancies; however it is the most common endocrine neoplasm comprising approximately 90% of all endocrine neoplasms (3, 4). Tumors arising from follicular epithelial cells exhibit a wide range of morphologic and behavioral features which have made the attempts to classify these tumors very challenging over the past 50 years. Molecular genetics has made significant advances in this area that indicate the need for a simplified classification of these common neoplasms. Most of the thyroid tumors, originate from the thyroid follicular epithelium (5). Papillary thyroid carcinoma (PTC) is the most common (~80%) of these, followed by follicular thyroid carcinoma (FTC) (~15%). Various genetic alterations in thyroid cancer have been unrevealed through recent molecular research, which have also been translated into clinical practice. The majority of genetic alterations observed in PTC and FTC are either point mutations (BRAF and RAS) or rearrangements (RET/PTC and PAX8/PPARγ) (5, 6). Point mutations of the BRAF gene are the most common molecular alterations observed in PTC. Among the BRAF mutations occurred in PTC the most common mutation involves nucleotide
1799, which results in a valine-to-glutamine substitution. These alterations are observed in 40-45% of all PTCs. These mutations lead to the constitutive activation of BRAF kinase. Constitutively activated BRAF kinase, in return, stimulates the MAPK pathway which plays a role in thyroid carcinogenesis (7). BRAF<sup>V600E</sup> mutation has been identified in neither follicular carcinomas nor benign entities, suggesting a specific role of this mutation for PTC. Hence, molecular testing for BRAF<sup>V600E</sup> mutation increases the diagnostic accuracy of cytological preparations of thyroid nodules (6, 8). One major drawback of BRAF mutation analysis in PTC is that BRAF mutation is typically detected in PTC with classical morphology and the tall cell variant. The rate of BRAF mutation is however low (10-15%) in the follicular variant of PTC, which creates the major diagnostic challenge in fine needle aspiration (FNA) cytology evaluation (Figures 1, 2). Although controversial, BRAF<sup>V600E</sup> has also been reported to be associated with parameters indicating aggressive tumor biology. Among these are tumor presentation at advanced stages, local extension of the tumor as well as metastatic disease at the time of diagnosis, which adds a prognostic utility to this mutation. Extrathyroidal extension has been shown to be associated with BRAF<sup>V600E</sup> mutations in PTC which are less than 2 cm in size (pT1) as well (7, 9-12). The K601E point mutation, small in-frame insertions or deletions around codon 600, and AKAP9/BRAF rearrangement, are the other rare BRAF mutation observed in PTC (13, 14).

The RET gene is first identified in 1985 by Takahashi as novel human transforming gene which is located on chromosome 10q11.2 (15). It encodes RET protein receptor tyrosine kinase which is expressed at high levels in thyroid parafollicular (C cells) cells. RET proto-oncogene can be activated through RET/PTC chromosomal rearrangement in follicular cells, which in return leads to the activation of MAPK signaling pathway (3). The majority of known RET rearrangements fuses the tyrosine kinase domain of RET to the 5’ portion of different genes. RET/PTC<sub>1</sub> and RET/PTC<sub>3</sub> are the most commonly observed RET/PTC rearrangements. RET/PTC<sub>1</sub> rearrangement is found in 60-70% of all RET/PTC-positive cases, whereas RET/PTC<sub>3</sub> is present in 20-30% (16-19). RET/PTC<sub>1</sub> rearrangement is
generally associated with classic variant of PTC, papillary growth, microcarcinomas, and benign behavior (2) (Figure 3). On the other hand, RET/PTC3 is associated with the solid variant of PTC, radiation associated tumors, and tumor aggressiveness. When appropriate techniques are used, detection of RET/PTC rearrangement is reported to be highly specific for PTC. Depending on the sample type, different detection techniques can be used. For samples such as FNA materials, fresh or snap-frozen tissue samples reverse transcriptase PCR can be the method of choice, whereas for formalin-fixed paraffin-embedded tissue samples fluorescence in situ hybridization is a better choice. RET/PTC-positive PTC typically present at a younger age, shows classic papillary morphology, and tends to metastasize to lymph nodes (10). RET/PTC rearrangements have also diagnostic utility for PTC in FNA samples since detection of clonal RET/PTC supports the diagnosis of PTC. This is especially useful for inadequate cytological preparations and for samples which have indeterminate cytology (20, 21). RET is reported to be overexpressed in medullary thyroid carcinoma (MTC) as well owing to the observation of RET protooncogene activation through germline mutations resulting in aberrant activation of the RET receptors in MEN2A, MEN2B, and familial MTC (3).

RAS gene family, which includes HRAS, KRAS, and NRAS, encodes intracellular G-proteins, leading to signal propagation from receptor tyrosine kinases and G-coupled receptors through different intracellular signaling pathways. Hence, mutational activation of RAS proto-oncogenes results in the activation of MAPK pathway. The NRAS codon 61 and HRAS codon 61 mutations are frequently observed in thyroid tumors, contrary to the frequent detection of
KRAS codon 12/13 mutations in most other types. The frequency of RAS mutations is higher in the follicular types as well as the poorly differentiated and anaplastic thyroid carcinomas. The frequency of RAS mutations in classical FTC are reported to be 40-50%, whereas the detection rate is around 20-40% in follicular adenomas (22-24). The majority of RAS mutations observed in PTC (10–20%) are detected in the follicular variant of this tumor (25) (Figure 4). It is also accepted that hyperplastic nodules showing clonal RAS mutations represent neoplasms and therefore should be classified as follicular adenomas.

The t(2;3)(q13;p25) translocation results in PAX8/PPARγ rearrangement (3). This rearrangement is detected in FTC with conventional histomorphology (30–40%) and to a lesser extent in oncocyctic variants (26, 27). PAX8/PPARγ-positive follicular carcinomas show certain clinicopathological features, i.e. younger age at presentation, smaller size, and more frequent vascular invasion. A small portion of follicular adenomas and follicular variant PTC also show this rearrangement (27, 28). Furthermore, it has been reported that follicular adenomas showing this rearrangement demonstrate similar immunohistochemical profile with their malignant counterparts, which indicates that these lesions are probably early lesions of FTCs (29).

The observation that Hurthle cell (oncocytic) carcinomas have less frequent PPARγ rearrangements indicates a separate molecular pathway from that of FTC.

In addition to the well known and more common genetic alterations, some very rare mutations are also detected in PTCs. One such mutation involves the TRK rearrangement (30) which is observed in less than 5% of PTC. Studies using array-based approaches in thyroid malignancies found out changes in gene expression profiles (31) which need further validation studies with larger number of samples (32).

MicroRNAs are noncoding, single-stranded RNA molecules which are 18- to 24-nucleotid long. They are involved in regulation of different cellular processes among which are cell death and proliferation. Differential expression of microRNAs has been reported in many human cancers, including endocrine neoplasms such as PTC, anaplastic thyroid carcinoma, FTC. Several independent studies suggest that microRNA may not have only functional but a diagnostic role in thyroid carcinoma as well. One such relationship is the reduced expression of c-kit in PTC due to its targeted downregulation by microRNA-221 and -222. Studies performed on cell lines harboring RET/PTC1 rearrangement revealed significant upregulation of four microRNAs; i.e. microRNA-128a, 128b, 139, and 200a and down-regulation of four other microRNAs; i.e. microRNA-154, 181a, 302b, 302c. Two cell lines harboring BRAFV600E point mutation showed significant upregulation of microRNA-200a, -200b and -141 with downregulation of microRNA-127, -130a and -144. Since these studies are performed on cell lines and a comparison to actual thyroid carcinoma tissues is not yet well established, it is yet not feasible to use these microRNA abnormalities for diagnostic and/or prognostic purposes (2).

**Hereditary Papillary Thyroid Carcinoma**

The familial PTCs are rare (33) and tend to be of the cribriform-morular variant of PTC. This variant of PTC occurs in the setting of familial adenomatous polyposis (FAP) presents predominantly in young women and can be either solitary or multiple. The biological behavior is similar to conventional PTC. The presentation of the cribriform-morular variant of PTC may precede the FAP diagnosis which suggests that these patients should be counseled to be tested for FAP (2).

**Figure 4**: Detection of NRAS mutation at codon 61 by pyrosequencing analysis in a follicular variant papillary thyroid carcinoma.
ADRENAL CORTEX

Traditionally, the diagnostic pathology of adrenal cortical diseases includes diffuse lesions and single or multiple nodular conditions. Adrenal cortical diseases are relatively rare but are associated with significant mortality and morbidity. Therefore, it is important to recognize and understand these diseases. In the recent years several new tools have become available for the purpose of better identifying different diseases and in addition to immunophenotyping, molecular markers have led to the definition of more accurate and reproducible categories of adrenal cortical neoplastic diseases.

Genetics of Adrenal Cortical Diseases

Adrenal Cortical Tumors in Familial Cancer Susceptibility Syndromes

Even if the majority of adrenal cortical carcinomas arise in a sporadic setting, a minority of cases are associated with familial cancer syndromes, including the autosomal dominant Li–Fraumeni and Beckwith–Wiedemann syndromes and, more rarely, the Gardner syndrome, multiple endocrine neoplasia type 1, neurofibromatosis type 1, and the Carney complex (34). The association of adrenal cortical carcinoma with two conditions that will subsequently be coded as the Li–Fraumeni and hereditary colon cancer syndromes had already been established in the 1980s.

Sporadic Adrenal Cortical Tumors

Gene Mutations

Germline alterations of tumor suppressor genes and oncogenes responsible for familial cancer syndromes have also been found as somatic alterations in sporadic adrenal cortical tumors, especially in carcinomas. Losses of the MEN1 gene locus at 11q13, but very infrequent gene mutations, have been detected in sporadic adrenal tumors (35, 36). Somatic mutations of the TP53 gene, as seen in Li–Fraumeni syndrome, as well as p53 protein accumulation can be detected in sporadic tumors. The detection of these changes are considered as signs of malignancy, being virtually absent in adenomas. Activation of the Wnt/β-catenin pathway as the result of CTNNB1 mutations has been documented in up to 40% of carcinomas, but also in a relevant proportion of adenomas (37, 38), especially nonsecreting and/or large-size tumors (39). Finally, somatic inactivating mutations or allelic losses of the PRKAR1A locus at 17q22–24, involved in the Carney complex, were also observed in sporadic adrenal cortex lesions (40). However, a significant proportion of adrenal cortical tumors lacks known genetic defects, and therefore, there are several studies ongoing to clarify molecular mechanisms alternative to gene mutations in the pathogenesis of these tumors.

Chromosomal Imbalances

A number of studies have reported that chromosomal aberrations are more frequent in malignant than in benign and hyperplastic adrenal cortical lesions. Gains, losses, and amplifications can be detected with either comparative genomic hybridization (CGH) or allelotyping techniques. In particular, gains in chromosomes 6q, 7q, 12q, and 19p, and losses in chromosomes 3, 8, 10p, 16q, 17q, and 19q, have been associated with a significantly worse survival of adrenal cortical cancer patients, which were independent of tumor size, tumor weight, and functional status of the tumor (41, 42). A strong relationship between tumor size and number of chromosomal aberrations was reported, with no gains or losses detectable in adenomas smaller than 5 cm; conversely, gains on chromosomes 4 and 5 and losses on 2, 11, and 17 were apparently restricted to carcinomas having a size of 7–20 cm. Overall, extensive genomic imbalances were detected in carcinomas using CGH technology. This suggests a complex molecular pathogenesis for the development of sporadic tumors which involves various genetic alterations for transformation and tumor progression (43, 44).

MicroRNA Profiling

Since the differential diagnosis of adenoma versus carcinoma is a challenging one in adrenal cortical lesions, several studies have attempted to find novel markers of malignancy including microRNA analysis. Among the frequently overexpressed or downregulated miRNAs, miR-483 (in both 3p and 5p isoforms) and miR-195 are those more consistently found overexpressed and downregulated, respectively, both at the tissue and serum levels. However, data concerning the prognostic role of these two miRNAs are controversial (45). miR-210 is another miRNA which was reported to be upregulated by different groups. It is the miRNA most consistently induced under hypoxia, and high levels were found associated with clinicopathological parameters of aggressiveness (necrosis and high Ki-67 proliferation index) and a poorer survival (46).

DNA Methylation Profiling

The role of DNA hypermethylation in adrenal cortical tumorigenesis has been evaluated in some recent studies. In addition to altered DNA methylation of the H19 promoter and the promoter methylation of TP53, a significant DNA hypermethylation of the RASSF1A promoter in adrenal cortical carcinoma, but not in adenoma, has been described,
suggesting an epigenetic mechanism for RASSF1A silencing in malignant adrenal cortical tumors. More recently, two clusters of adrenal cortical carcinomas based on CpG island methylation status, the CpG island methylator phenotype ("CIMP") and "non-CIMP" were identified, the former associated to a poorer prognosis (45).

PARATHYROID

Parathyroid pathology which often gives rise to hyperpara-thyroidism (HPT), can be due to adenoma, hyperplasia or carcinoma. In the vast majority of cases (80-85%) HPT is primary and is the result of an adenoma which is a disease of one of the parathyroid glands. The histopathological and molecular discrimination of adenoma (PA) versus carcinoma (PT-CA) can create a real challenge in routine pathology practice. This topic has been discussed in corresponding review article in this special issue.

GASTROINTESTINAL TRACT

Gastrointestinal Neuroendocrine Tumors (GI NET)

Gastrointestinal neuroendocrine tumors (GI NETs) are a group of tumors which arise from diffuse neuroendocrine system cells. Common to this group of tumors is the expression of neuroendocrine markers and production of certain peptide or amine hormones.

Molecular Genetics

Since GI NETs are a heterogenous group of tumors, the molecular mechanisms involved in these tumors are also heterogenous and poorly understood. Several studies have shown that genes common in many other tumor types are not involved in the pathogenesis of GI NETs which is also the case for pancreatic endocrine neoplasms (PEN). Among the limited number of genes involved in GI NET development, mutations of MEN1 gene have often been observed in association with endocrine tumors of the stomach, duodenum, and pancreas (47). Mutation of the NFI gene, on the other hand, leads to an increased risk for periampullary tumors (48). Several studies have revealed frequent alterations of chromosome 18q, which contains important genes such as Smad2, Smad4 and DCC genes. Another important observation is the detection of LOH in X-chromosome or 17q13 only in malignant NETs. The mutations of Reg1 alpha gene is associated with histamine producing ECL cell tumors in the setting of hypergastrinemia (49).

One of the most common genetic alterations observed in PENs, following MEN1 mutations, is the LOH on chromosome 3p, which has been detected in around 30% of the cases. Studies have failed to demonstrate significant roles for tumor suppressor genes or oncogenes in PENs which are commonly altered in other tumor types. Array-based techniques revealed alterations of several different group of genes which may play an important role not only in the pathogenesis but in the prognosis of PENs as well (50-53).

Among these are oncogenes, growth factor –related genes, cell adhesion and migration molecules, potential biomarkers, therapeutic targets, cell cycle checkpoint genes, transcription factors and apoptosis –related genes (54-56). However one major drawback of these observations is the low level of concordance among the published data, which requires further investigations in larger PEN sample groups.

ACKNOWLEDGEMENTS

We thank Arda Gunay and Aybuke Kabaoglu, MSc for their technical assistance and Sehbal Arslankoz, MD for her valuable help during preparation of the manuscript.

CONFLICT OF INTEREST

The authors have declared no conflict of interest.

REFERENCES


